

Sustained Cell Quality in Cryopreserved Leukopaks: Viability, Phenotype, and Expansion Potential

Lisa Paschold¹, Julia Uhlig², Norbert Lidzba², Irene Gómez-Gràcia³, Katharina Renate Winterstein¹, Anke Wahler¹, Barbara Ingrid Baumann-Baretti¹, Francisco J Belda³, Anna Dünkel², Daniel Gonnermann¹

¹Haema Bio Supplies Division, Haema GmbH, Leipzig, Germany; ²Fraunhofer Institute for Cell Therapy and Immunology (IZI), Leipzig, Germany; ³Grifols, Bio Supplies, Research and Development Department, Barcelona, Spain

BACKGROUND

- ❖ Cryopreservation of leukopaks (Fig. 1) is essential for flexible manufacturing.
- ❖ Its impact on cell quality remains critical for cell-based therapies¹⁻³.
- ❖ Current characterization of frozen leukopaks primarily relies on cell viability and recovery.



FIGURE 1. Leukopak.

AIM

- ❖ Investigate the impact of cryopreservation on PBMC composition and T cell characteristics.

METHODOLOGY

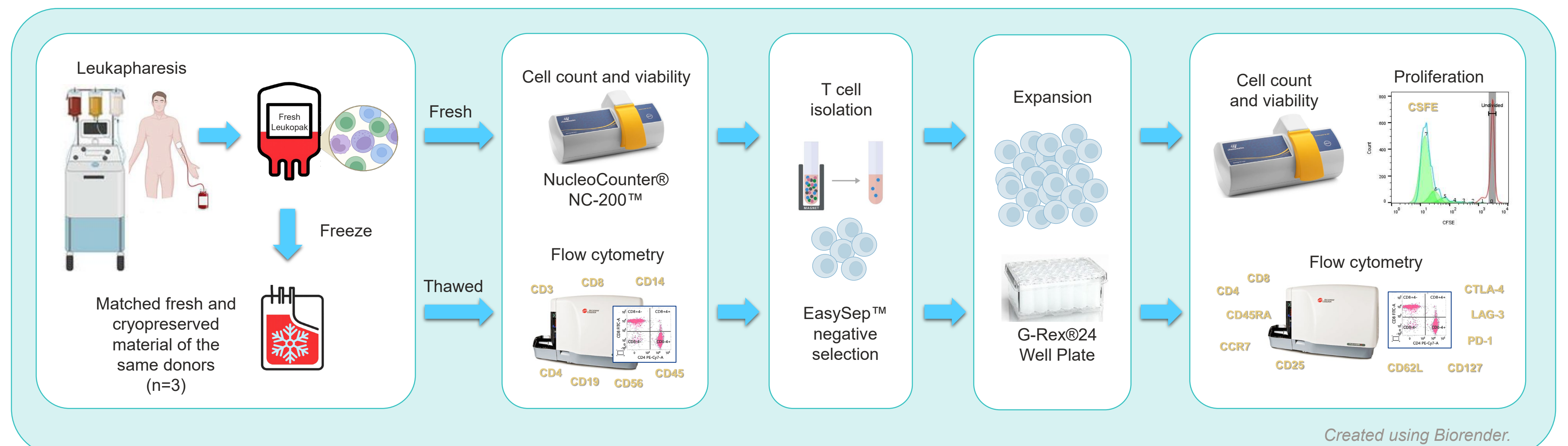


FIGURE 2. Experimental workflow.

RESULTS

The overall PBMC composition was comparable between fresh and cryopreserved material (Fig. 3A). While T cell isolation yielded comparable purity across conditions (Fig. 3B), recovery was reduced in frozen samples, likely due to diminished CD3 expression following thawing.

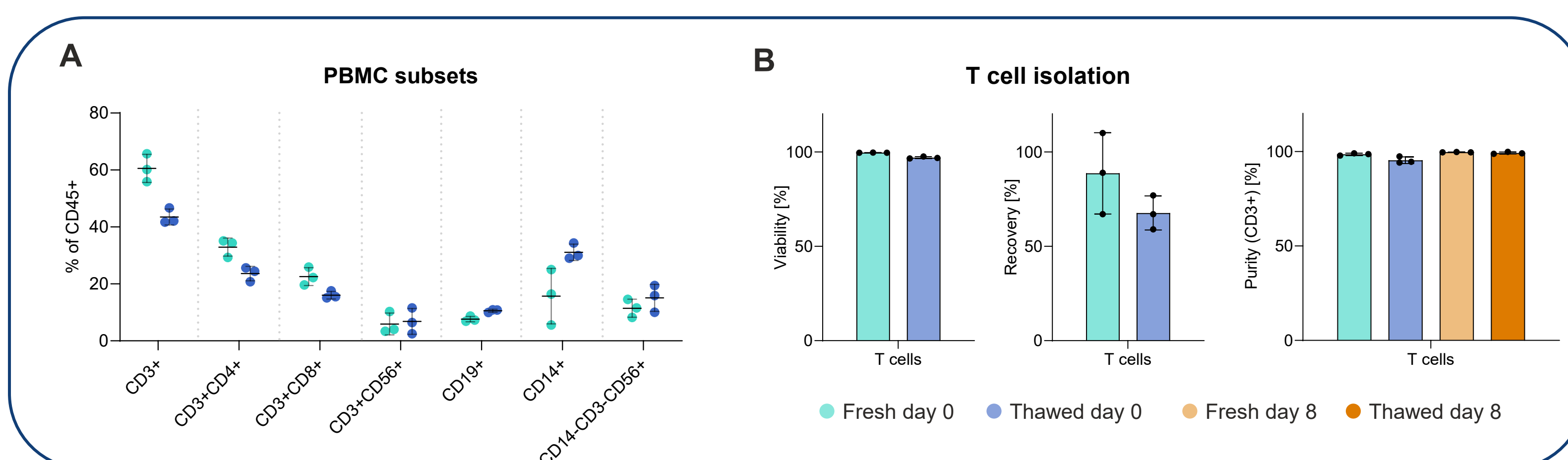


FIGURE 3. PBMC phenotype and T cell isolation in fresh vs. thawed leukopak material. (A) PBMC surface markers of leukopak material. (B) Viability, recovery and purity after T cell isolation.

Post-thaw viability of T cells remained high after thaw, cell isolation and cultivation (>94%) (Fig. 4A). Cryopreservation did not affect viability (Fig. 4A), CD4/CD8 ratios (Fig. 4B), expansion (Fig. 4C) or proliferation (Fig. 4D) after culture for up to 8 days.

Activation and exhaustion markers remained low and were not induced by freeze-thaw (Fig. 5).

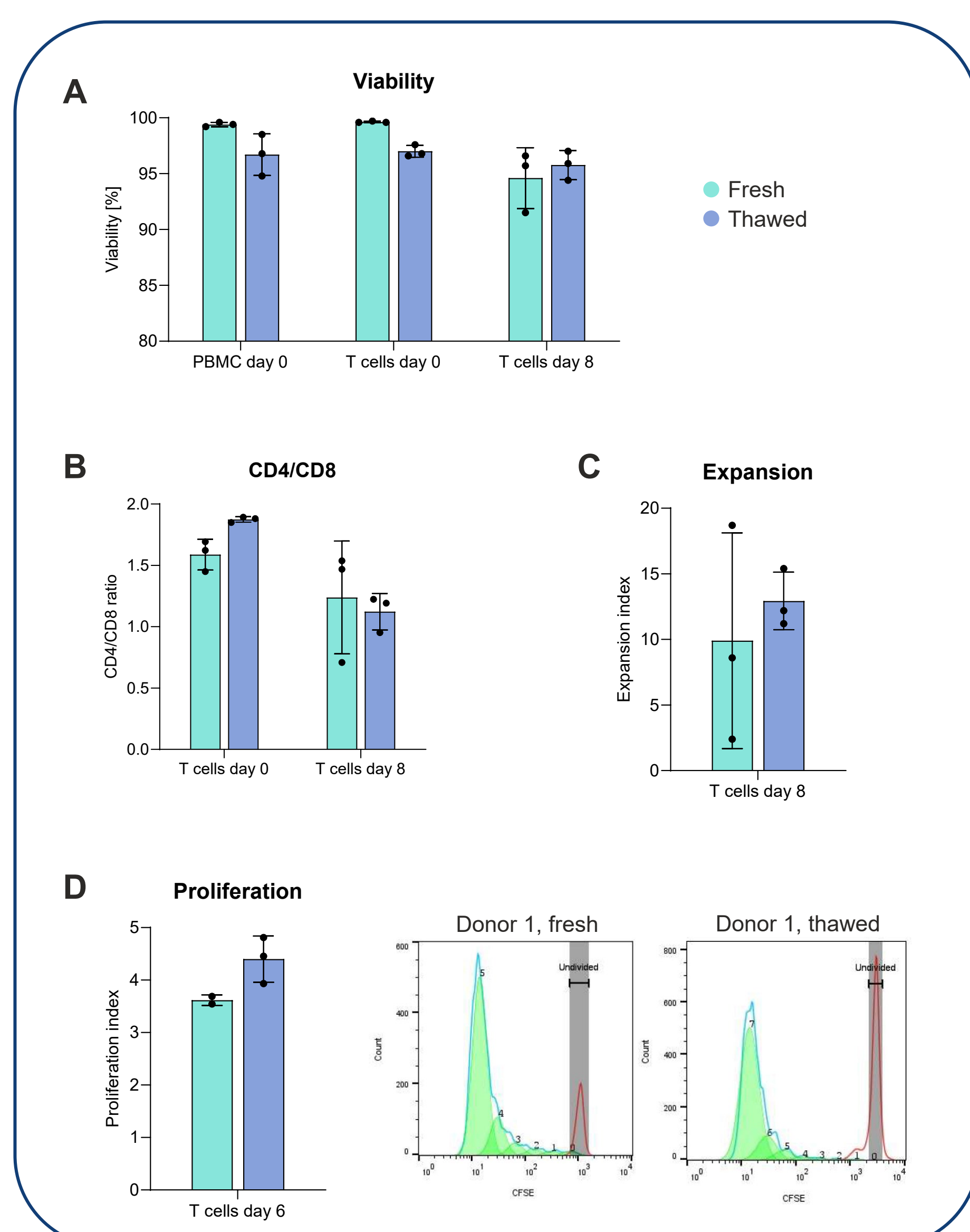


FIGURE 4. Expansion and proliferation of T cells isolated from fresh vs. thawed leukopak material. (A) Viability of PBMCs and T cells. (B) CD4/CD8 ratio. (C) Expansion index determined by cell count. (D) Proliferation index determined by CFSE staining and exemplary deconvolution plots.

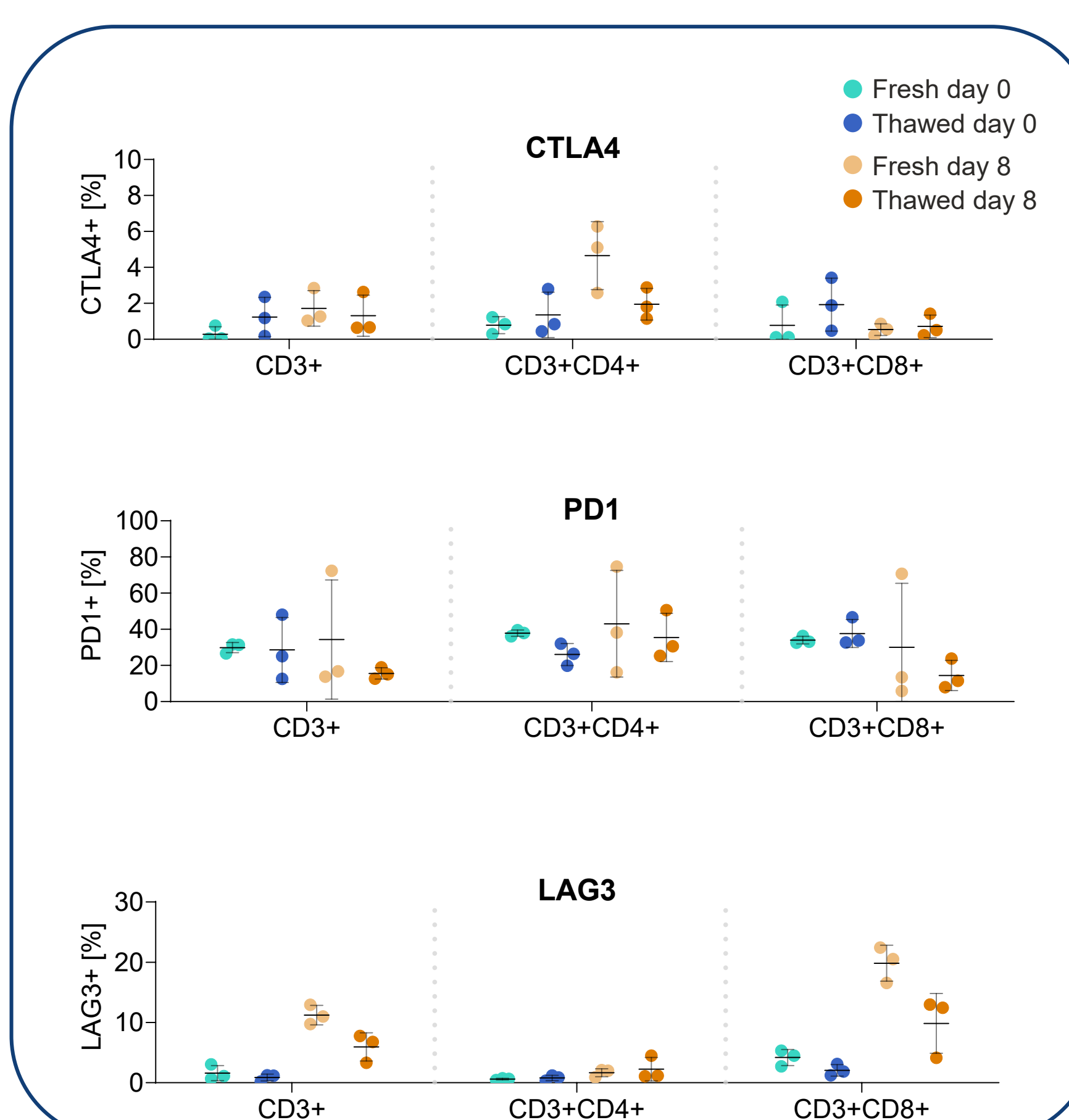


FIGURE 5. Activation status of T cells isolated from fresh vs. thawed leukopak material. CD3+, CD3+CD4+ or CD3+CD8+ T cells directly after cell isolation and after 8 days of culture were further analyzed for markers of activation and exhaustion using flow cytometry.

CONCLUSION

- ❖ Cryopreservation maintained viability, CD4/CD8 ratios, proliferation, and expansion.
- ❖ Minor memory shifts post-thaw normalized after culture.
- ❖ Activation/exhaustion markers remained low throughout.
- ❖ Our cryopreservation protocol preserves key CAR T-relevant T cell attributes, throughout freeze-thaw processing.

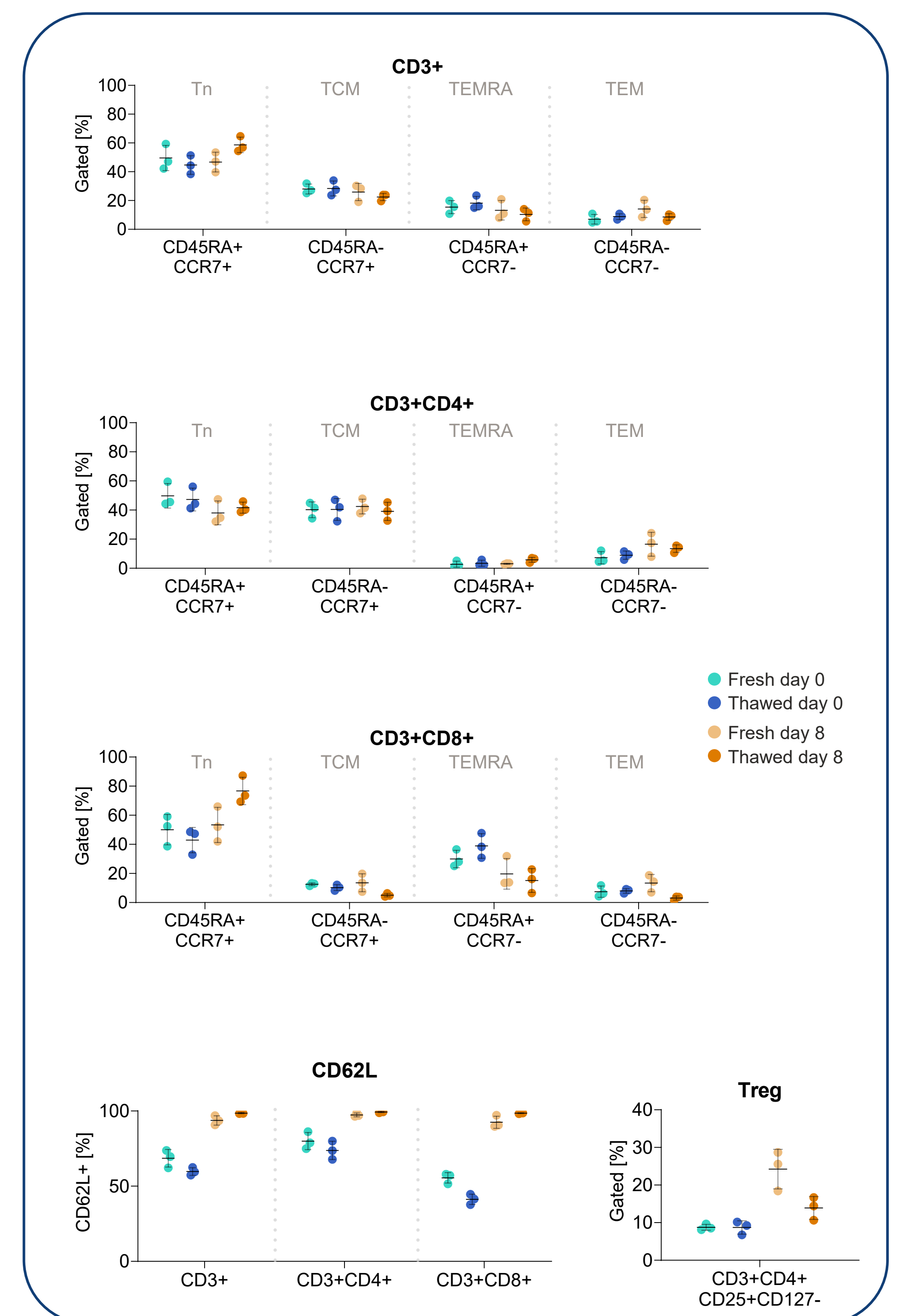


FIGURE 6. Cell compartments of T cells isolated from fresh vs. thawed leukopak material. CD3+, CD3+CD4+ or CD3+CD8+ T cells directly after cell isolation and after 8 days of culture were further analyzed for markers of memory compartments and Tregs using flow cytometry.

Minor shifts in memory subsets were observed immediately post-thaw but normalized after culture (Fig. 6). T_{reg} levels were preserved post-thaw but declined during expansion, indicating selective outgrowth of effector T cells.

Based on the results of this study, further investigations are planned as part of the SaxoCell collaboration.

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REFERENCES

- 1 Tyagarajan, S. et al., *Cytotherapy*, 2019; 21(12):1198-1205.
- 2 Eastwood, G. et al., *JITC*, 2015; 3:P383.
- 3 Ren, M. et al., *Sci Rep*, 2025; 29870.

